

Network Motifs

Simple Building Blocks of Complex Networks

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13.12.2016

outline

introduction

genetics

motifs

introduction

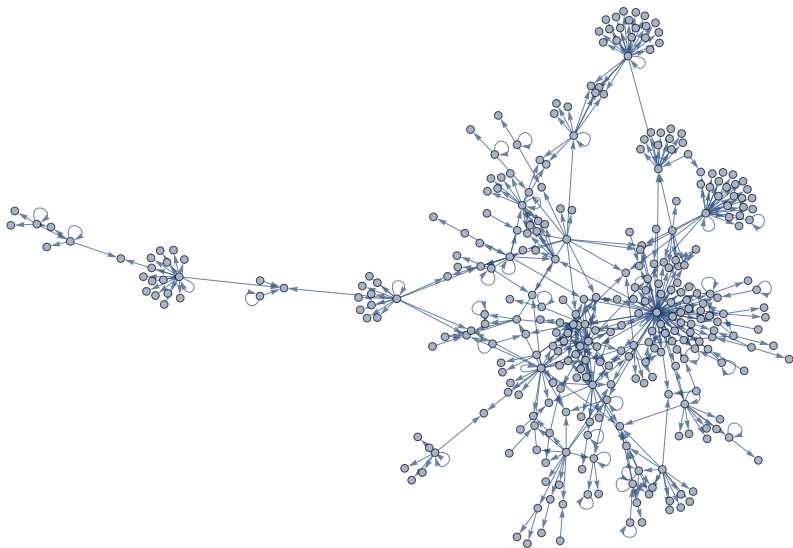
motivation

methods

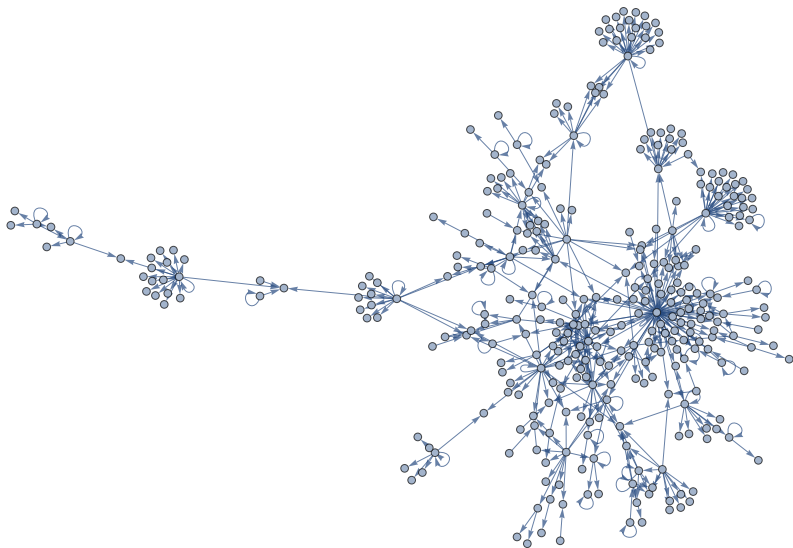
genetics

motifs

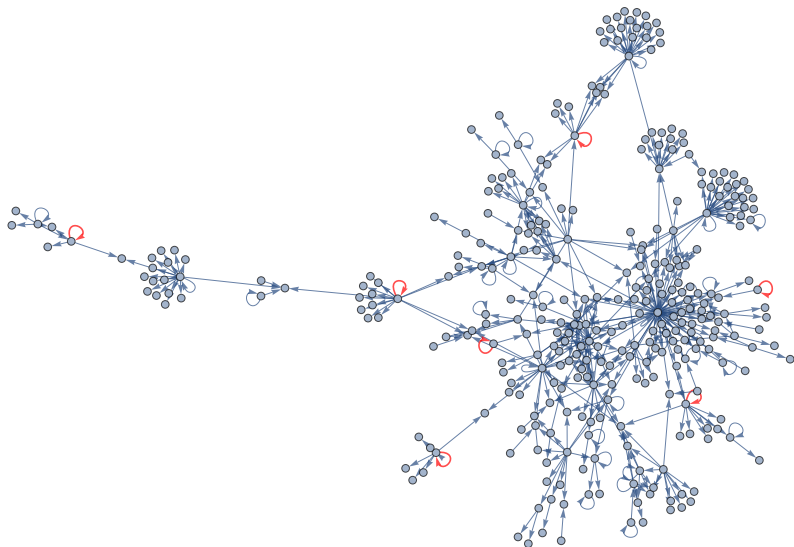
fraction of transcription network of *E. coli*



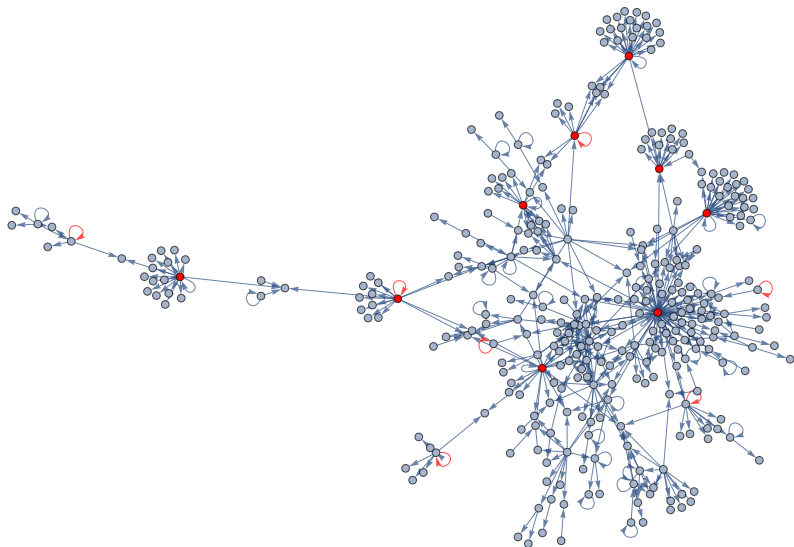
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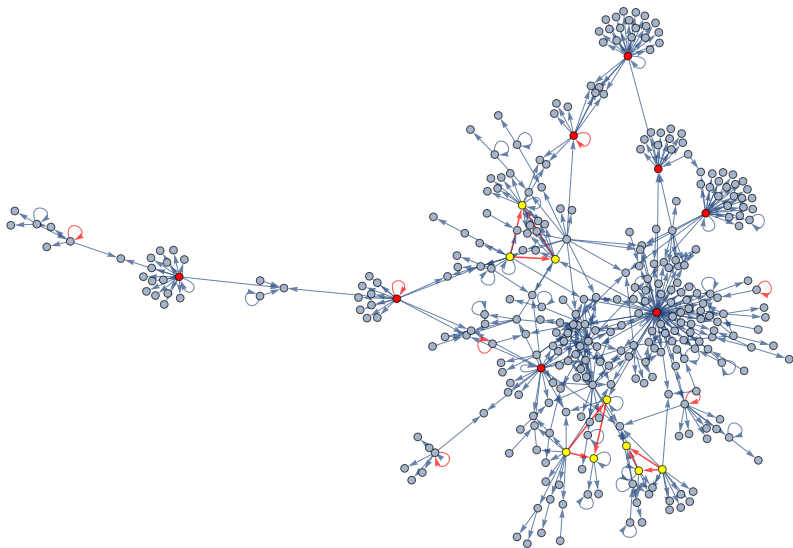
transcription network - autoregulation



transcription network - outstanding nodes



transcription network - loops



Is there a statistically significant aggregation of local patterns or "motifs" in the network?

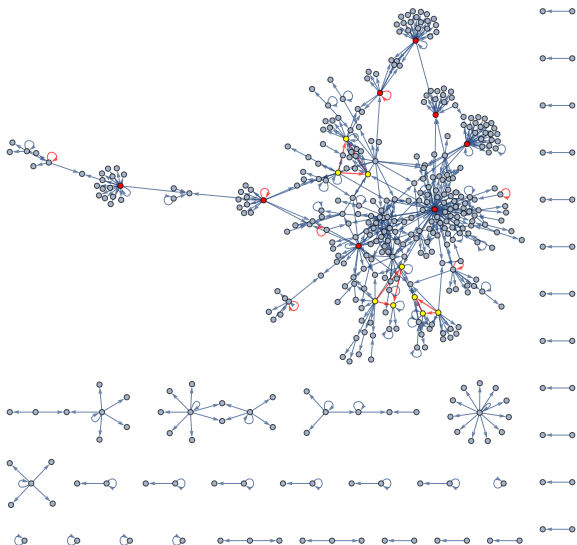
Compare to a randomized network

→ YES!

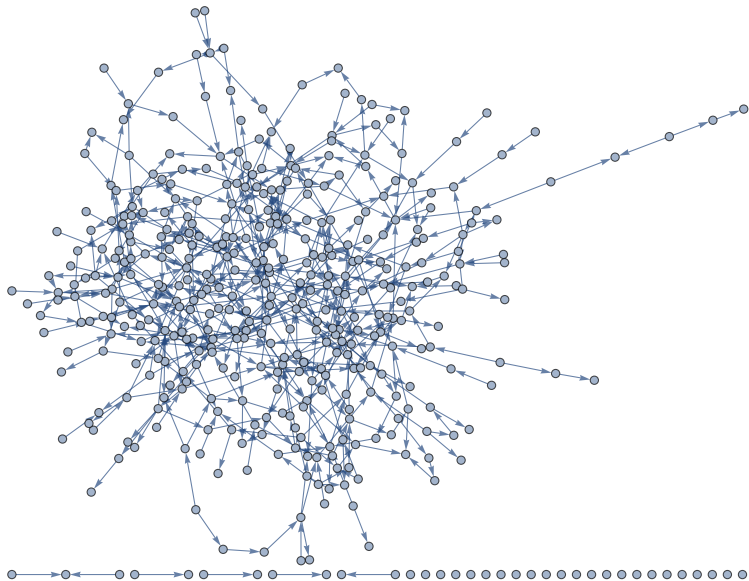
Which are the motifs that evolution prefers?

What are their functions in the network?

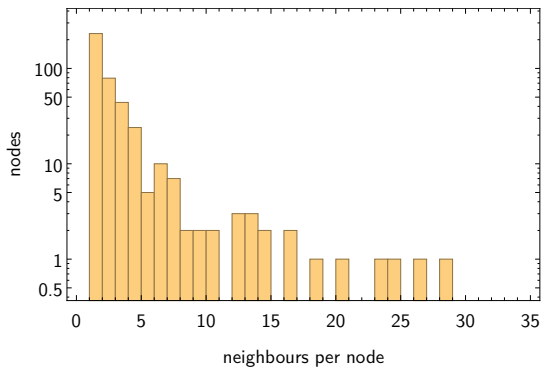
fraction of transcription network of *E. coli*



random network



histogram of E.coli transcription network



Randomization: Choose two random edges, swap target nodes
 → the histogram is preserved



introduction

genetics

transcription

translation

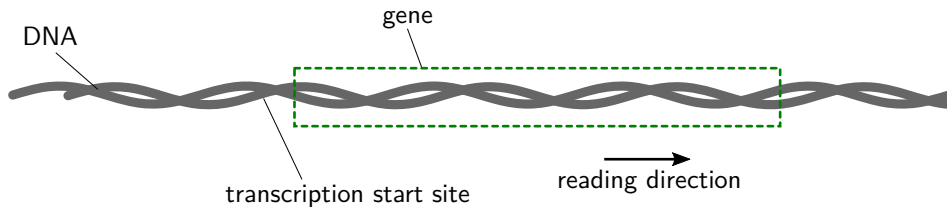
dynamics

motifs

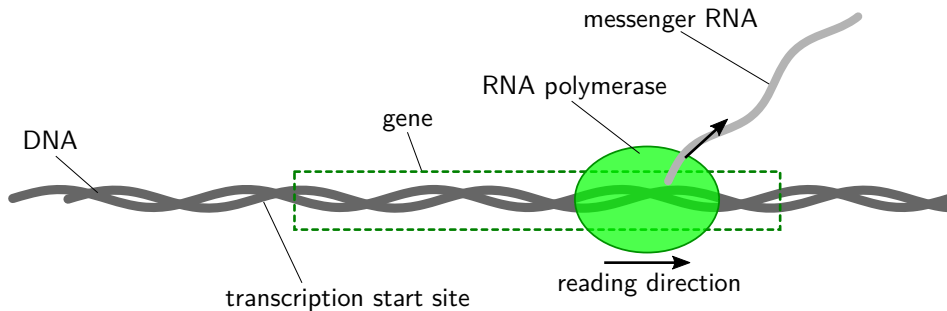
gene expression - transcription



gene expression - transcription

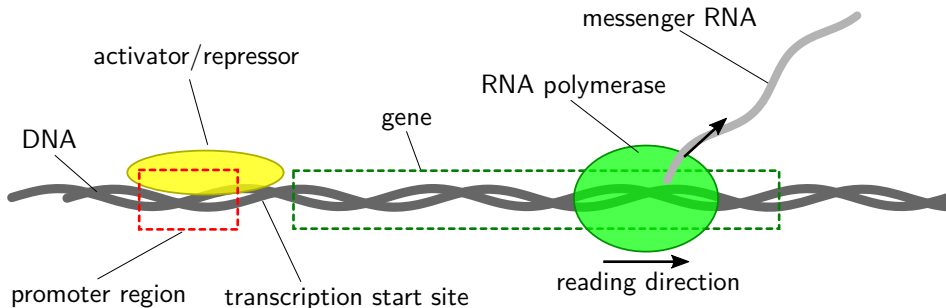


gene expression - transcription



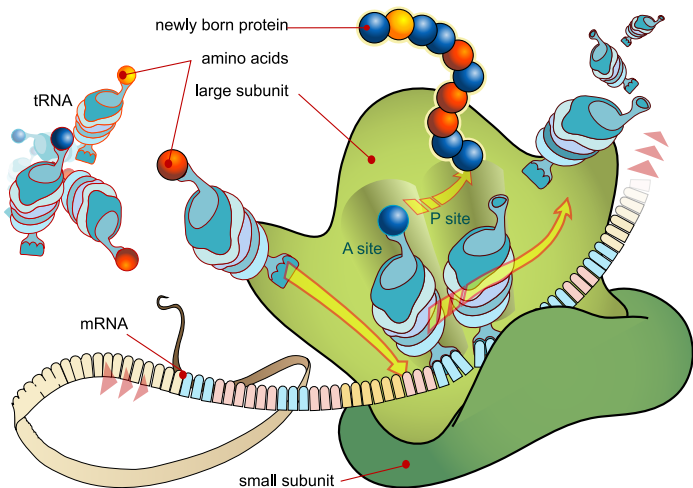
- RNAP transcribes DNA to mRNA

gene expression - transcription



- RNAP transcribes DNA to mRNA
- Activator/repressor interacts with transcription start site
→ Enhances/inhibits attachment of RNAP

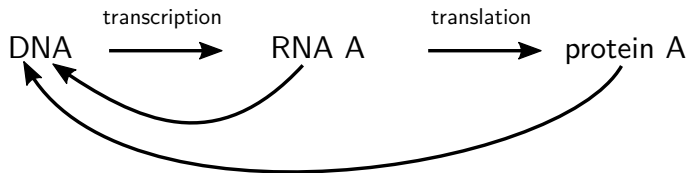
gene expression - translation of mRNA to protein



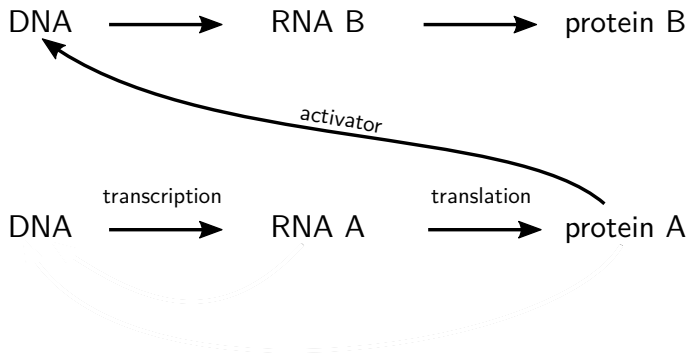
regulation of gene expression



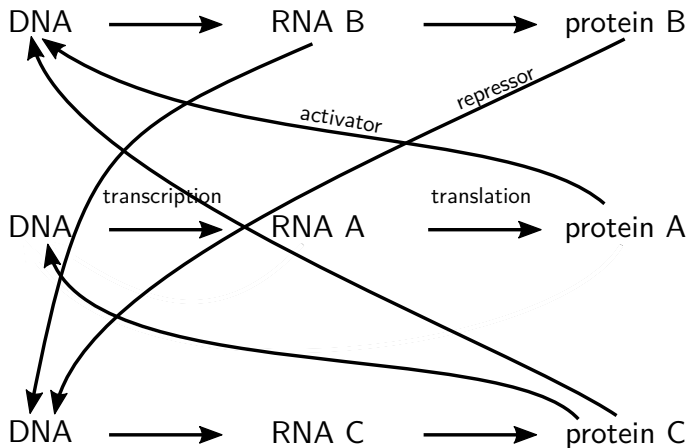
regulation of gene expression



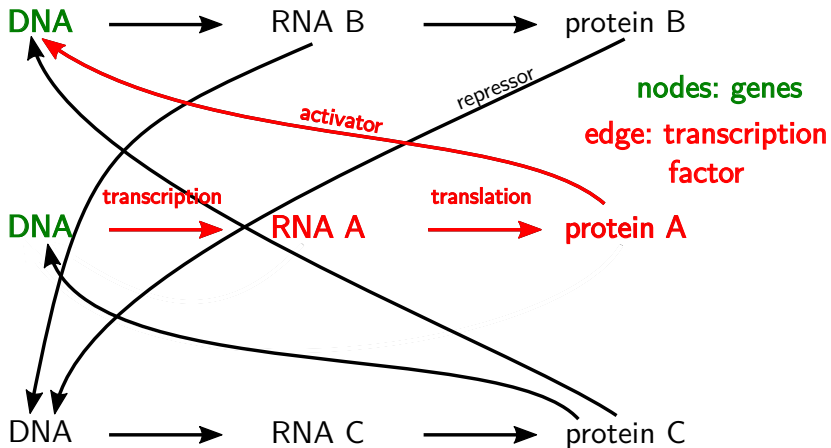
regulation of gene expression



regulation of gene expression



regulation of gene expression



dynamics of simple gene regulation

Transcription factor X regulates expression of protein Y: $X \rightarrow Y$

Simple assumptions:

- If X is in active form X^* , Y is produced at a constant rate β
- Degradation rate of Y is α

$$\Rightarrow \frac{dY}{dt} = \beta - \alpha Y$$

$$\text{Stable state: } \frac{dY}{dt} \stackrel{!}{=} 0 \quad \Rightarrow \quad Y_{\text{st}} = \frac{\beta}{\alpha}$$

dynamics of simple gene regulation

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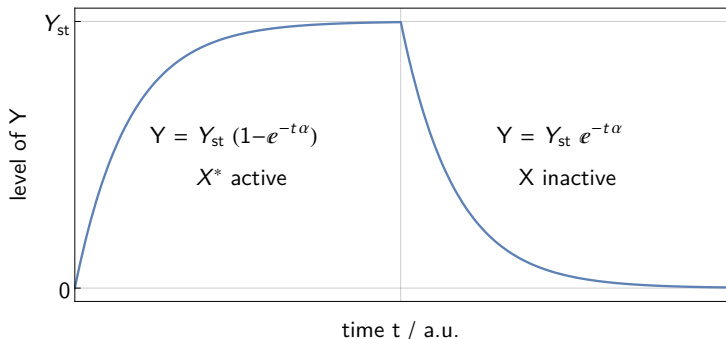
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dynamics of simple gene regulation

$$\frac{dY}{dt} = \beta - \alpha Y$$

$$Y_{st} = \frac{\beta}{\alpha}$$



Hill-function

More realistic model: Rate of production of Y is a function of X^*
 $f[X^*]$ has three parameters \rightarrow each edge carries three numbers

- K : Level of X^* to significantly activate expression
- β : Maximal expression level
- n : Cooperativity: Number of molecules needed for activation

$$\text{Rate of production of Y: } f[X^*] = \beta \frac{X^{*n}}{K^n + X^{*n}}$$

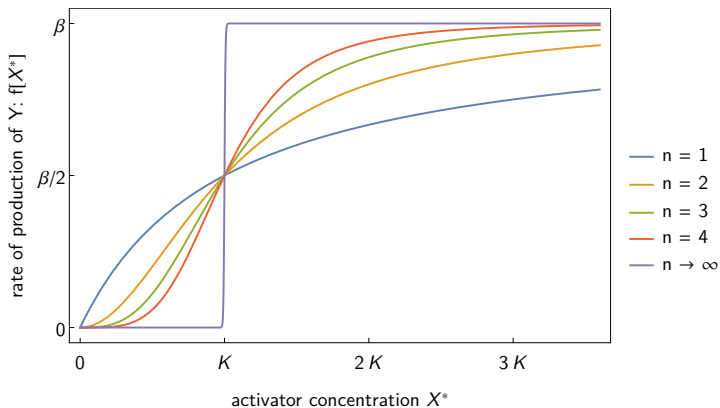
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Hill-function



$$f[X^*] = \beta \frac{X^{*n}}{K^n + X^{*n}}$$

Hill-function

Parameters can be tuned during evolution:

K : Mutations of binding site in the promoter area

β : Mutations in the RNAP binding site

$$\text{Rate of production of Y: } f[X^*] = \beta \frac{X^{*n}}{K^n + X^{*n}}$$

introduction

genetics

motifs

- autoregulation

- feed forward loop

- single-input module

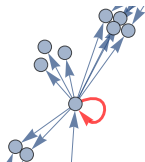
- dense overlapping regulon

- occurrence of motifs in various networks

autoregulation

Probability of a self-edge in random network with N nodes:

$$p_{\text{self}} = \frac{1}{N}$$



Probability of k self-edges in a random network with N nodes and E edges:

$$P(k) = \binom{E}{k} \cdot p_{\text{self}}^k \cdot (1 - p_{\text{self}})^{E-k}$$

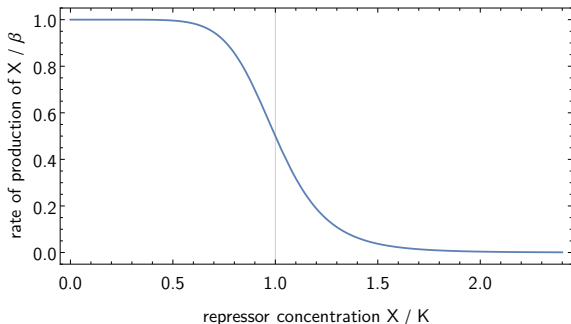
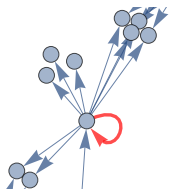
The expectation value for the number of self-edges is

$$\langle N_{\text{self}} \rangle = E \cdot p_{\text{self}} = \frac{E}{N}$$

For the E.coli network this yields $\frac{E}{N} = \frac{519}{424} = 1.2$ self-edges

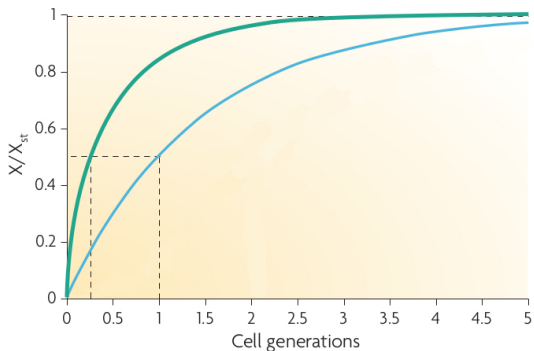
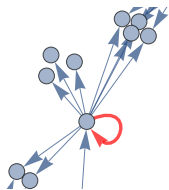
The real network has 40 self-edges \rightarrow high statistical significance

negative autoregulation



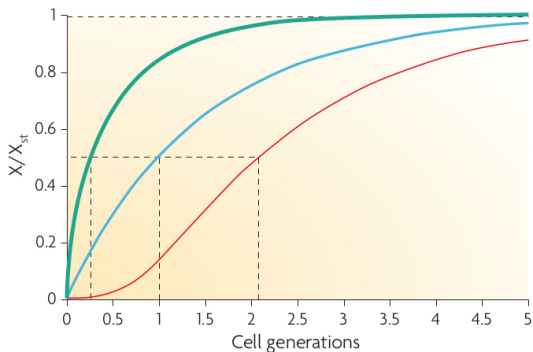
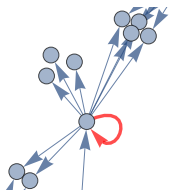
- System with simple regulation: $Y_{st} = \frac{\beta}{\alpha}$
 → unstable, many factors influence β and α
- System with negative autoregulation: $Y_{st} \approx K$
 → stable, K is specified by strength of chemical bonds
 ⇒ Noise suppression

negative autoregulation



- High production rate β can cause strong initial rise
 - High autorepression leads to saturation at stable level K
- \Rightarrow response acceleration

positive autoregulation



— Negative autoregulation
 — Simple regulation
 — Positive autoregulation

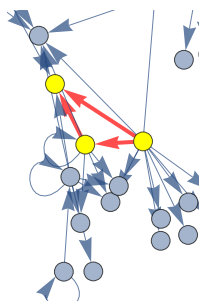
⇒ higher response time

⇒ noise amplifying

Why is noise good for a cell?

→ diversity

feed forward loops



Coherent FFL

Coherent
type 1



Coherent
type 2



Coherent
type 3



Coherent
type 4



Incoherent FFL

Incoherent
type 1



Incoherent
type 2



Incoherent
type 3



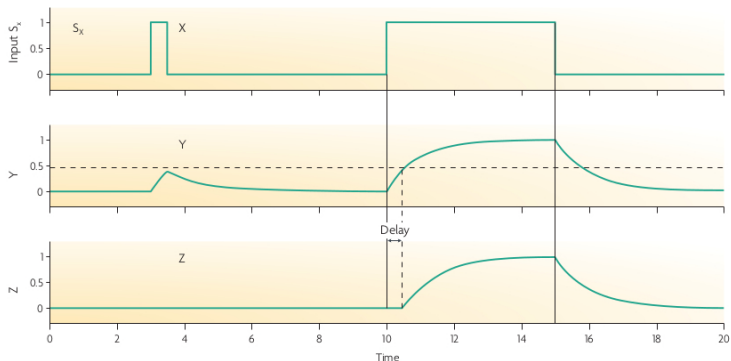
Incoherent
type 4



coherent type 1 feed forward loop

Coherent FFL

Coherent
type 1



AND connection for Z

→ only respond to persistent stimulation, elevator door effect

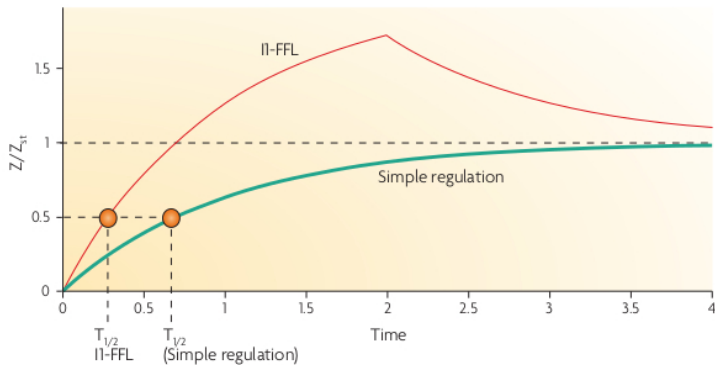
OR connection for Z

→ no delay after stimulation, but delay after stimulation stops

incoherent type 1 feed forward loop

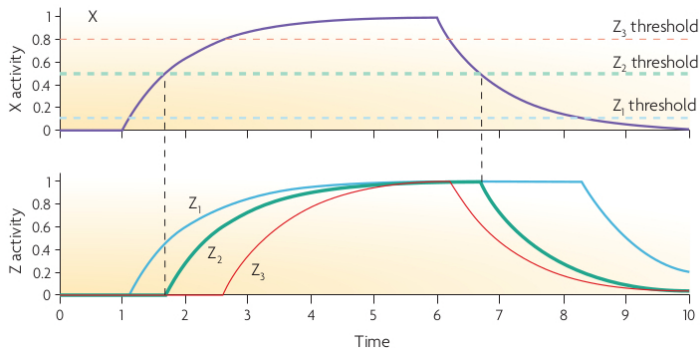
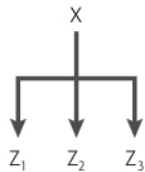
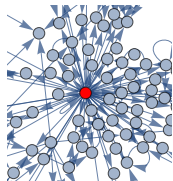
Incoherent FFL

Incoherent
type 1



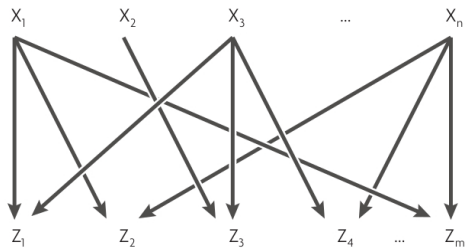
- Response acceleration
- Pulse generator

single-input module



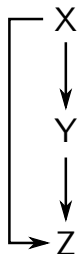
→ timed expression of different genes

dense overlapping regulon



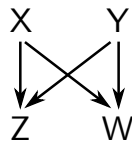
- Not yet very well understood
- Detailed information about connection strength is needed

motifs in the transcription network of E. coli



feed-forward loop

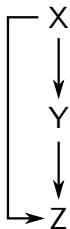
$$N_{\text{real}} = 40 \quad N_{\text{rand}} = 7 \pm 3$$



bi-fan

$$N_{\text{real}} = 203 \quad N_{\text{rand}} = 47 \pm 12$$

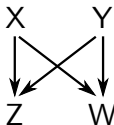
motifs in the neural network of *C. elegans*



feed-forward loop

$$N_{\text{real}} = 125$$

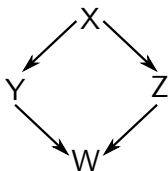
$$N_{\text{rand}} = 90 \pm 10$$



bi-fan

$$N_{\text{real}} = 127$$

$$N_{\text{rand}} = 55 \pm 13$$



bi-parallel

$$N_{\text{real}} = 227$$

$$N_{\text{rand}} = 35 \pm 10$$

motifs in the food web of little rock

X

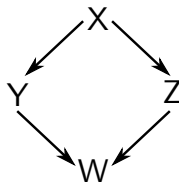


Y



Z

three chain

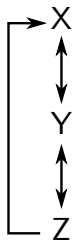


bi-parallel

$$N_{\text{real}} = 3219 \quad N_{\text{rand}} = 3120 \pm 50$$

$$N_{\text{real}} = 7295 \quad N_{\text{rand}} = 2220 \pm 210$$

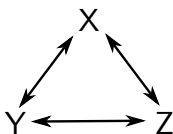
motifs in the world wide web



feedback with two
mutual dyads

$$N_{\text{real}} = 110000$$

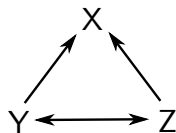
$$N_{\text{rand}} = 2000 \pm 100$$



fully connected triad

$$N_{\text{real}} = 6800000$$

$$N_{\text{rand}} = 50000 \pm 400$$

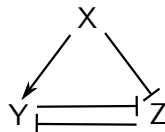
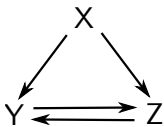


uplinked mutual dyad

$$N_{\text{real}} = 1200000$$

$$N_{\text{rand}} = 10000 \pm 200$$

motifs developmental networks



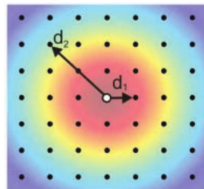
a careful look at the statistical methods

What has been done?

- Real network $\xrightarrow{\text{randomization}}$ random network
 - Posing of null hypothesis:
“The occurrence of motifs is the same in both networks“
 - The null hypothesis is rejected by a statistical test
- Conclusion:
Evolution prefers motifs that are overrepresented and disfavours others

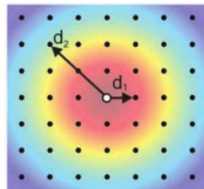
a careful look at the statistical methods

- Create a random *toy model*
- Probability of connection between two nodes reduces with distance
- Test model against random network with same # of nodes and edges
- There is a significant occurrence of motifs in the toy model!
 - It is clearly not evolution which prefers those motifs
 - Spatial clustering can create motifs



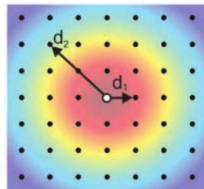
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references

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- Shen-Orr, S. S., Milo, R., Mangan, S. & Alon, U. *Network motifs in the transcriptional regulation network of Escherichia coli*. Nature Genet. 31, 64–68 (2002).
- Milo, R. et al. *Network motifs: simple building blocks of complex networks*. Science 298, 824–827 (2002).
- Alon, U. *Introduction to Systems Biology: Design Principles Of Biological Circuits* (CRC, Boca Raton, 2007).
- Artzy-Randrup, Y. et al. *Comment on “Network Motifs: Simple Building Blocks of Complex Networks” and “Superfamilies of Evolved and Designed Networks”*. Science 305, 1107 (2004).

résumé

- We can identify repeated small patterns in real networks
- Comparison to randomized networks shows a significant accumulation of motifs in real networks
- We have to be careful which random network to use as null hypothesis
 - We can describe the behaviour of the isolated motif

